

Journal Pre-proof

Fine powder of lipid microparticles – spray drying process development and optimization

Eliza Wolska

PII: S1773-2247(21)00320-8

DOI: <https://doi.org/10.1016/j.jddst.2021.102640>

Reference: JDDST 102640

To appear in: *Journal of Drug Delivery Science and Technology*

Received Date: 21 February 2021

Revised Date: 19 May 2021

Accepted Date: 31 May 2021

Please cite this article as: E. Wolska, Fine powder of lipid microparticles – spray drying process development and optimization, *Journal of Drug Delivery Science and Technology*, <https://doi.org/10.1016/j.jddst.2021.102640>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

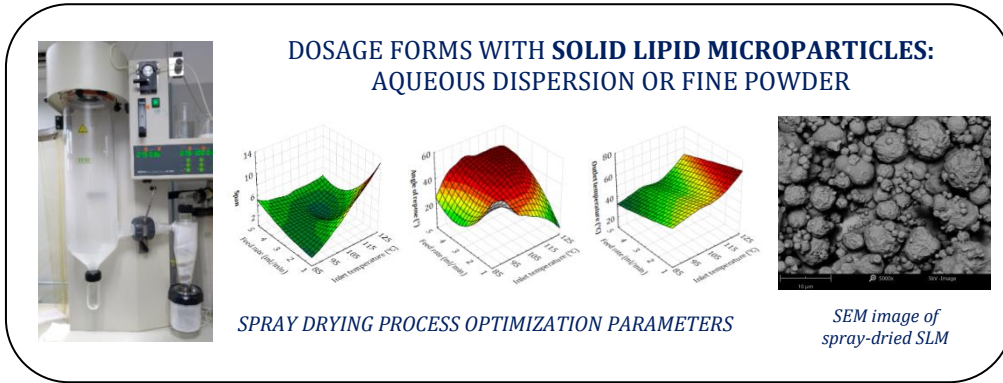
© 2021 Published by Elsevier B.V.



Author Contributions:

Eliza Wolska: conceptualization, methodology, investigation, writing

Journal Pre-proof



Journal Pre-proof

1 Fine powder of lipid microparticles – spray drying process 2 development and optimization

3
4 Eliza Wolska

5 *Department of Pharmaceutical Technology, Medical University of Gdansk, Hallera 107, 80-416*
6 *Gdansk, Poland*

7 * *Corresponding author:* Tel. +48 58 3491085; fax: +48 58 3491090.

8 *E-mail address:* eliw@gumed.edu.pl (Eliza Wolska)

10 Abstract

11 **Objectives:** Performed studies were focused on developing spray drying technique for
12 aqueous dispersion of solid lipid microparticles (SLM) by selecting appropriate process
13 parameters and assessing their impact on the process and properties of the obtained dry SLM
14 powders.

15 **Significance:** Spray drying allows to obtain SLM in a dry powder form when the liquid form
16 does not present sufficient long-term stability (e.g. due to degradation of the active substance
17 or aggregation of particles) or when the dosage form is to be used in a fine powder form.

18 **Methods:** In the first stage of research the experiments were designed to optimize process
19 parameters during spray drying of the *placebo* SLM dispersions prepared with two lipids:
20 Compritol or stearic acid. The inlet temperature and feed rate were process parameters
21 selected for monitoring. As response values, yield and quality attributes of the final product,
22 namely particle size, moisture content and powder flowability were chosen. The process
23 parameters optimized in the first step were then used to dry the SLM with model active
24 substances: cyclosporine and spironolactone.

25 **Results:** The use of 3D surface charts, developed on the basis of the results of the conducted
26 experiments, allowed for the selection of optimal process conditions for obtaining final
27 product with desired properties and satisfying yield. For SLM with Compritol these were:
28 inlet temperature 90°C and feed rate 2.4 ml/min; whereas for SLM with stearic acid 80°C and
29 3 ml/min were optimal, respectively. Process parameters optimized for *placebo* formulations
30 were found to be equally suitable for drying drug-loaded SLM.

31 **Conclusions:** The spray drying was found to be an effective method of obtaining dry powders
32 from aqueous SLM dispersions. The lipid forming the SLM matrix should be considered the
33 most important factor on which the process parameters depend. The most appropriate drying
34 conditions selected during drying *placebo* formulations proved to be equally effective when
35 SLM with the same composition and with model active substance were subjected to drying.

36 *Keywords:* solid lipid microparticles, spray drying, microspheres, cyclosporine,
37 spironolactone, powder flowability

38

39 **1. Introduction**

40 Solid lipid microparticles (SLM) are lipid-based formulation with great potential as
41 drug delivery system. SLM were developed on the basis of SLN (solid lipid nanoparticles)
42 studies, but they are in the micrometers size range (usually 1-100 μm , up to 1000 μm ,
43 depending on the preparation method [1-4]). Due to the particle size and the solid state of the
44 lipid forming the matrix, SLM can provide prolonged drug release. In comparison to SLN
45 higher drug loading is also feasible [5-8]. Similar like for SLN, the most common techniques
46 for the preparation of SLM dispersions are: melt dispersion technique (called also hot
47 emulsification method), solvent evaporation or diffusion method and microchannel
48 emulsification technique [4]. Although SLM present beneficial technological and
49 biopharmaceutical properties, they are incomparably less studied as drug carriers than SLN
50 [9-11]. As biocompatible multicompartiment carrier SLM are a good alternative to polymer
51 microparticles and are considered for both, systemic (oral, parenteral [12-14]) and topical
52 (dermal, ocular or even inhaled [15-18]) application.

53 SLM can be applied as a liquid dispersion (aqueous suspension of microspheres) or a
54 fine powder (e.g. inhalation powders) depending on the intended administration route [3, 18,
55 19]. Producing SLM in the form of a dry powder is also justified in order to increase the long-
56 term stability of the formulation (physical, chemical and microbiological) [20, 21]. Although
57 the long-term stability studies confirmed that SLN and SLM formulations in the liquid
58 suspension form remained stable after 2 years of storage [8, 22, 23], there is a greater risk of
59 adverse changes in the aqueous suspensions, especially in drug loaded preparations.
60 Unfavorable physicochemical transformations may concern both the lipid matrix and other
61 excipients, as well as the active substance (API), which may undergo chemical degradation
62 (e.g. hydrolysis) or premature release. Changes like degradation of the particles matrix,
63 particle aggregation/particle fusion or unwanted increase in particle size were observed in
64 both SLM and SLN dispersions [3, 20]. After conversion into dry powders, lipospheres could
65 be stored over a long period, without the risk of physicochemical changes characteristic for
66 liquid dispersions. Obtained dry formulations could be used in the form of a powder or after
67 reconstitution, as a suspension. It is essential that the reconstituted dispersion exhibits the
68 same properties as the original suspension.

69 There are few and not very well studied methods for producing SLM in the form of a
70 dry powder: spray congealing (also called spray chilling), spray drying (from organic
71 solution), cryogenic micronisation or particles from gas-saturated solutions technique (PGSS)
72 [4]. However, the main problems reported are: large size of the obtained lipid particles (even
73 up to 2000 μm) and/or the use of organic solvents (e.g. ethanol) [4, 24]. A solution to both
74 these issues can be a two-stage process: 1/ hot emulsification technique for preparation of
75 SLM aqueous dispersion and 2/ evaporation of water by spray drying of the resulting
76 suspension. While it is known that spray drying process may convert various liquid feed, not
77 only solution, but also suspension or emulsion to a dried particulate form, this approach was
78 only used by Mezzena et al. [17] for the production of inhalable SLM from microparticulate
79 dispersion. In other published reports, the lipid microparticles were obtained by spray drying
80 of organic solutions – lipids were dissolved in dichloromethane, chloroform or ethanol [15,
81 25, 26]. The spray-drying process should certainly be considered as a more favorable
82 alternative to the more expensive and time-consuming lyophilization process, which could
83 also be applied as a method of transforming an aqueous dispersion into dry powder. In
84 addition, during freeze-drying, lipospheres are exposed to freezing and desiccation stress,
85 which may be detrimental to their further stability. Several sugars (glucose, fructose,
86 trehalose, and sorbitol) are being used as cryoprotectants to overcome that concerns since they
87 have shown ability to conserve lipid carrier properties after freeze-drying. Different sugars
88 could have different cryoprotectant power at concentrations used (usually in the range from
89 5% to 10%), providing various protection to the SLN or SLM during freeze-drying and
90 storage.

91 Drying is a one-step process often used for conversion of a liquid formulation into a
92 dry powder. Spray drying is a simple, fast and scalable technology used widely not only in
93 pharmaceutical but also in food and chemical industries [4, 27]. The fluid is atomized into
94 thermal contact with a hot drying medium (usually air) with a temperature usually up to
95 220°C, depending on the properties of the material to be dried. Although the sprayed liquid is
96 in contact with a hot gas, it occurs for a short time and the cooling effect of the evaporating
97 solvent conserves the droplet temperature relatively low. Thus, even heat-sensitive products
98 can be dried with a negligible degradation. It is also possible to dry lipid systems, such as
99 SLN or SLM, even at temperatures above 100°C. SLN dispersions were successfully dried at
100 the inlet temperature of 110°C [18, 19]. However, because this temperature is higher than the
101 melting point of the lipid forming the liposphere matrix, in case of improperly selected
102 process parameters, one should expect the phenomenon of partial melting during the process.

103 This could result in a change, or even a significant deterioration of the properties (mainly
104 related to powder flow, but also API distribution and the drug release behavior) of the
105 obtained lipospheres.

106 The yield of the spray drying process and properties of the obtained powders are
107 directly influenced by process parameters. Several factors are known to affect the spray
108 drying process, e.g. inlet temperature, feed rate, outlet temperature or concentration of solids
109 in the feed [4, 27]. With regard to lipid particle dispersions the influence of temperature
110 during drying is especially important. The particles delivered in feed may also be adversely
111 affected by high shear stress in the nozzle. In effect it is not so easy to maintain the size, shape
112 and morphology of the primary particles, which is the main goal of process optimization.
113 Although the spray drying process exhibits many applications in pharmaceutical industry, the
114 use of this process for conversion of the aqueous dispersion of SLM into a dry powder is not
115 sufficiently understood or well reported.

116 The aim of the present study was to examine and determine on a laboratory scale the
117 best instrument parameters and working conditions to transform aqueous SLM dispersion into
118 SLM dry powder, not losing primary size and morphology in such a way that the obtained
119 SLM dry product can further be used directly as a dry powder or after reconstitution into a
120 liquid dispersion before administration. A practical aspect of the work was to identify the
121 most critical process and formulation parameters in order to achieve successful performance
122 of the process yielding a product with the desired particle size and flowability. As drying of
123 the drug-loaded formulation may not be as effective as for the *placebo* formulation, in the
124 next step dispersions of SLM loaded with model APIs (cyclosporine – CsA and
125 spironolactone – SPIR) were spray dried using the process parameters optimized during the
126 first stage. To the best of our knowledge this is the first research paper presenting the
127 optimization method of spray drying process of an aqueous SLM dispersion with a particle
128 size less than 50 μm .

129

130 **2. Materials and methods**

131 *2.1. Materials*

132 Cyclosporine A (CsA) was obtained from LC Laboratories (Boston, MA, USA) and
133 Compritol 888 ATO (glyceryl behenate) from Gattefossé (Saint-Priest, France).
134 Spironolactone (SPIR), stearic acid and Tween 80 (polysorbate 80) were purchased from

135 Sigma-Aldrich (St. Louis, MO, USA); polyvinylpyrrolidone (PVP) was from BASF
136 (Ludwigshafen, Germany). All other chemicals used were of analytical reagent grade.

137

138 2.2. Preparation of SLM dispersions

139 For the preparation of SLM dispersions two different lipids: Compritol and stearic acid
140 were used as matrix-forming lipid. The lipid concentration in the dispersions was 10% (w/w)
141 and the formulations were either prepared drug-free (*placebo* formulations) or drug-loaded
142 with cyclosporine (F/CsA) or spironolactone (F/SPIR). The composition of all tested
143 formulations was selected at the preliminary stage of SLM dispersion examination and is
144 illustrated in Table 1. The drug loaded formulations contained 0.1% or 1.0% (w/w) of CsA
145 and 0.1% or 0.5% (w/w) of SPIR, which in relation to the lipid content was 1% or 10% of
146 CsA and 1% or 5% SPIR, respectively. The experimentally determined solubility of the active
147 substances in the tested lipids was 100 mg/g (10%) and 330 mg/g (33%) of CsA in Compritol
148 and stearic acid, respectively, as well as about 30 mg/g (3%) of SPIR in both tested lipids. In
149 comparison with selected API concentration in SLM formulations, SPIR concentrations were
150 both below (1%) and above (5%) the specified solubility, while in the case of CsA were
151 below (1%) or equal to the determined solubility (10%).

152 SLM formulations were prepared using a hot emulsification method, which has been
153 fully described in a previous paper [16]. All excipients were heated at a temperature 10°C
154 above the lipid melting point, which was 69-74°C for glyceryl behenate and 69,6°C for stearic
155 acid [28, 29]. Thereafter, the mixing process of the lipid phase with aqueous phase was
156 performed at 80°C using a high-shear mixer Ultra-Turrax (T25 Janke-Kunkel, IKA
157 Labortechnik, Germany) with dispersing tool from stainless steel (S 25N – 18 G, working
158 range from 10 ml to 1.5 L), at the speed of 8000 rpm for 5 min. After cooling in an ice bath
159 (30 min) the dispersions were stored in a refrigerator. The batch size was 200 g or 300 g.

160

161

162

163

164

165

166

167

168 **Table 1**169 The composition (w/w %) of the investigated formulations *placebo* and with API

Formulation	The composition of formulations (<i>placebo</i> and with API)					
	CsA	SPIR	Compritol	Stearic acid	Tween 80	Water
F1	-	-	10.0	-	5.0	85.0
F2	-	-	-	10.0	3.0	87.0
F3	0.1	-	10.0	-	5.0	84.9
F4	1.0	-	10.0	-	5.0	84.0
F5	0.1	-	-	10.0	3.0	86.9
F6	1.0	-	-	10.0	3.0	86.0
F7	-	0.1	10.0	-	5.0	84.9
F8	-	0.5	10.0	-	5.0	84.5
F9	-	0.1	-	10.0	3.0	86.9
F10	-	0.5	-	10.0	3.0	86.5

170

171

172 *2.3. Spray drying of SLM dispersions*

173 Spray drying of tested formulations was performed using a laboratory Buchi Mini
 174 Spray Dryer B-290 (Buchi Labortechnik AG, Flawil, Switzerland) equipped with standard 0.7
 175 mm nozzle. SLM dispersions immediately before spray drying were diluted with 5% (w/w)
 176 PVP solution in equal parts (1:1) and stirring on a magnetic stirrer until the drying process
 177 was completed. In the main stage of the study, *placebo* SLM were tested (Table 1). The
 178 instrument parameters such as: inlet temperature and liquid feed rate were changed within the
 179 range indicated in Table 2. The outlet temperature was evaluated because its value depended
 180 on both of these parameters. Other process parameters were maintained constant: the pressure
 181 of compressed air was 0.7 MPa, aspiration setting was 100% and the air was used as a drying
 182 gas. As response values, to determine efficiency of the process and quality of the final
 183 product, particle size (span and percentage of particles with size <50 μm), the yield, moisture
 184 content and flowability of the SLM powders were examined (Table 2).

185

186

187

188

189

190

191

192 **Table 2**

193 Ranges in which the instrument parameters were modified and the properties of the spray
 194 dried SLM powders which were evaluated

Tested factors	Range of instrument parameters		Investigated powder properties
	Compritol	Stearic acid	
Inlet temperature	90-120°C	75-100°C	Particle size (µm)
Feed rate	1.2-6.0 ml/min	2.4-4.5 ml/min	Span
Outlet temperature*	32-62°C	32-50°C	Yield (%)
* a value that was not set at the beginning of the process, but resulted from the other two: inlet temperature and feed rate			Moisture content (%)
			Flow rate (min)
			Angle of repose (°)
			Hausner ratio (HR)

195

196

197 The resulting spray dried powders were collected and stored in capped glass jars at
 198 room temperature. Tested parameters were modified based on the properties of the obtained
 199 product. The important feature assessed was the process efficiency calculated as the process
 200 yield according to Eq. (1).

$$201 \quad \text{Yield (\%)} = \frac{\text{Practical yield}}{\text{Theoretical yield}} \times 100 \quad (1)$$

202 where *practical yield* is the amount of powder recovered after spray drying in the receptacle
 203 and *theoretical yield* is the amount of dry mass from the dispersion without losses.

204 In the spray dried powders moisture content, particle size and span (see point 2.4. and
 205 2.6.) were also evaluated. Flow properties of the prepared dry SLM powders were tested as
 206 well (see point 2.7.). In Table 2 both equipment-related factors, as well as investigated
 207 properties of tested formulations were presented. During the process optimization, attempts
 208 were made to select drying parameters that could guarantee the best properties of the obtained
 209 dry powders. As the best properties of lipid particles (shape, morphology, size) were
 210 considered properties as close as possible to the primary particles (from SLM dispersion).
 211 Due to the different possibilities of further use of the spray dried SLM powder, its flowability
 212 was also assessed, following the pharmacopoeial criteria for determining the angle of repose
 213 and the Hausner ratio.

214 In the second stage of research, the inlet temperature and flow rate parameters, which
 215 have already been selected as the most appropriate for *placebo* formulations, were applied

216 during API loaded SLM (Table 1) spray drying. As a model drugs CsA and SPIR were
217 chosen. In this work, the obtained API loaded dry powders were also characterized as
218 described above. In this case, however, in addition to the physicochemical properties, such as
219 in the case of placebo particles (shape, size, flowability), it is also important to assess the
220 impact of spray-drying process on the biopharmaceutical properties of lipid particles.
221 Therefore, the distribution of the active substance in the lipid matrix or the release rate of API
222 from the dosage form has been described in detail in a separate paper [30].

223

224 2.4. Particle size analysis

225 The particle size distribution was measured by laser diffraction (Beckman-Coulter LS
226 13 320, Indianapolis, IN, USA). When the particles were in aqueous SLM dispersion (after
227 preparation, before spray drying) Universal Liquid Module (ULM) was used. ULM is capable
228 of measuring particle samples in the size range 0.017 μm to 2000 μm due to additional
229 detectors and PIDS function (*Polarization Intensity Differential Scattering*). SLM dispersion
230 was added to the sample cell until the correct obscuration parameter (usually at the level of
231 40-45%) was obtained (when sizing particles with using PIDS, a PIDS obscuration level of
232 40% to 60% is recommended).

233 Spray dried powders were also measured by laser diffraction, this time using a
234 Tornado Dry Powder System (DPS) connected to the same device (Beckman-Coulter LS
235 13 320) as the ULM attachment. The measurement of powder was carried out without
236 dispersion in a liquid medium and without any other sample preparation prior to
237 measurement. The appropriate amount of sample was placed in a sample holder and delivered
238 to the sensing zone in the optical bench by a vacuum. The Tornado DPS provides automatic
239 feed rate (obscuration) control to obtain the set point for the obscuration (6%). DPS is capable
240 of measuring particle samples in the size range 0.4 μm to 2000 μm .

241 The obtained results were recorded in the form of a graphs and statistics presenting
242 values e.g. d_{10} , d_{50} , d_{90} , determined as measures of maximum diameter of 10%, 50% and 90%
243 of the detected particles, respectively. Particle size was also expressed as span calculated
244 using the following Eq. (2).

$$245 \quad \text{Span} = \frac{d_{90} - d_{10}}{d_{50}} \quad (2)$$

246 Finally, the value corresponding to the percentage of particles located below the value of
247 50 μm in the tested formulation was also evaluated.

248

249 *2.5. Optical microscopy and scanning electron microscopy*

250 An optical microscope (Nikon, Eclipse 50i, Nikon Corporation, Tokyo, Japan) was
251 used for initial microscopic evaluation of *placebo* and drug-loaded SLM.

252 To visualize surface properties and morphology of the tested lipid microparticles a
253 scanning electron microscope Phenom Pro (Phenom World Thermo Fisher, Eindhoven,
254 Netherlands) was employed. Standard sample holder and a carbon adhesive tape were used to
255 fix a sample of SLM dry powder. When the SLM dispersion was tested, water was evaporated
256 from the sample at room conditions after the dispersion was applied to the carbon tape. Before
257 microscopic observations tested sample was coated with a thin layer of gold. Acceleration
258 voltage of 5 kV was applied to record images at a magnification of 5000x.

259

260 *2.6. Moisture content determination*

261 The moisture content of the spray dried SLM powders were analyzed using Moisture
262 Analyzer type WPS 210S (Radwag, Poland). The appropriate mass of the powder samples
263 were uniformly spread to a thin layer on a sample dish and dried to a constant weight with a
264 heating cycle of up to 105°C. It was considered that at this temperature no degradation of any
265 compound would occur but that all the water would be evaporated. The weight loss after
266 complete drying reflects the initial and therefore total moisture content of the tested samples.

267

268 *2.7. Flowability assessment*

269 Flow properties of the prepared spray dried SLM powders were tested according to
270 pharmacopoeial methods (Ph. Eur. 9.0). The flow rate and angle of repose were measured
271 using a manual powder flow tester (Electrolab, Mumbai, India). A powder samples were
272 placed in a funnel with 10 mm orifice and poured onto the base with diameter (d) of 10 cm to
273 form a cone. The height of the cone (h) was measured using height gauge and the angle of
274 repose α ($^\circ$) was calculated using following Eq. (3).

$$275 \quad \tan(\alpha) = \frac{h}{0.5 \times d} \quad (3)$$

276 Measurements were done in triplicate, and the results were expressed as mean \pm standard
277 deviation (SD). As reported by the pharmacopoeia, although there is some variation in the
278 qualitative description of powder flow using the angle of repose, much of the pharmaceutical

279 literature appears to be consistent with the classification by Carr, which is shown in European
280 Pharmacopoeia and was used to classify our powders.

281 Hausner ratio (HR) was determined with the procedure in which the unsettled apparent
282 volume (V_o) and the final tapped volume (V_f), of the powder, after tapping the material until
283 no further volume changes occur, were measured and HR was calculated according to Eq. (4).

$$284 \quad \text{Hausner Ratio} = \frac{V_o}{V_f} \quad (4)$$

285 The tapped volume was determined using Erweka SVM tester (Heusenstamm, Germany), by
286 measuring the volume of the powders after 1250 taps. The results of the Hausner ratio were
287 classified according to the generally accepted scale of flowability presented in the European
288 Pharmacopoeia (2.9.36. Powder flow).

289

290 2.8. Statistical analysis

291 All the statistical analyzes were conducted and all data charts were prepared using
292 Statistica software (StatSoft program, Version 12). The statistical significance of differences
293 between assessed factors was tested by a one-way analysis of variance ANOVA. Differences
294 were considered to be significant at level of $p < 0.05$.

295

296 3. Results and discussion

297 3.1. Selection of conditions for the spray drying process

298 The spray drying process is one-step method to transfer SLM from aqueous dispersion
299 to a form of dry powder. This method requires the careful adjustment of drying conditions
300 appropriate to the material being dried in order to obtain a product with the desired properties.
301 This is particularly important for primary dispersion composed of lipid material with low
302 melting point. Therefore, in our study, optimization of spray drying conditions of SLM
303 aqueous dispersions was performed, and the aim was to obtain SLM powder composed of
304 lipid particles, the properties of which remained unchanged after the spray-drying process.

305 The first attempts to spray-dry the aqueous SLM dispersions indicated numerous
306 difficulties in obtaining SLM in the form of a dry powder with the desired properties. Due to
307 the lack of literature reports (which concern mainly aqueous or organic solutions of active
308 substances and polymers, and not lipid suspensions), the process was carried out under
309 various conditions of inlet temperature, flow rate or concentration of the dried dispersion

310 (2.5%, 5%, 10%). The problem turned out to be not only to obtain a fine, dry powder (that can
311 also be redispersed in water), but also the melting of the solid lipid during drying or to poorly
312 yield of the process. Polymers (polyvinylpyrrolidone – PVP, hydroxypropylmethylcellulose
313 and maltodextrins) were used as auxiliary substances, not only facilitating the drying process
314 but also redispersion. The polymers in the composition of the dried mixture are also used for
315 coating or obtaining a prolonged action. Polymers are known, that can act as an agent
316 avoiding particle aggregation, stabilizing agent and filler [4, 31]. Among the tested polymers,
317 the best properties were shown by PVP, which was selected for further tests as an auxiliary
318 substance facilitating drying and subsequent redispersion.

319 A number of the observed problems related to the drying of the SLM dispersion
320 indicated the need for a precise selection of the process conditions, and thus the need for its
321 optimization. For this purpose, *placebo* SLM dispersions with Compritol or stearic acid were
322 prepared according to the composition in Table 1. The PVP solution was added to the SLM
323 suspensions in a 1:1 ratio prior to spray drying. Then aqueous dispersions were dried
324 according to the process conditions shown in Table 2. Inlet temperature was investigated from
325 75 to 120°C and feed rate from 1.2 to 6.0 ml/min, with specific narrowing for SLM with
326 stearic acid. At the same time, influence of outlet temperature on the powder particles features
327 was investigated. The outlet temperature depends on inlet temperature and flow rate
328 simultaneously, and is a factor that could be used to control the course of the entire process.
329 The initial selection of drying process parameters and their experimental range presented in
330 Table 2 were based on the formulation properties (lipids melting point), our previous
331 experience and observations carried out during preliminary studies. Based on the results of
332 preliminary experiments the primary dispersions contain 5% (w/w) and that value was
333 maintained constant in the current studies, because the drying process also depends on the
334 SLM concentration.

335 In dry SLM powders, the particle size distribution was examined, and the moisture
336 content, yield and flowability were determined. Characterization of SLM powders was
337 conducted at room conditions. The type of analyzed physicochemical properties of the tested
338 powders was selected on the basis of their potential impact on the application properties, long
339 term stability and process properties, if they were to undergo further technological processes.

340

341 *3.2. Spray drying of aqueous dispersions of Compritol-SLM*

342 For all obtained 13 batches of *placebo* SLM dispersions with Compritol (F1 in Table
343 1) the process was feasible in the assumed parameters ranges of the apparatus (Table 2),
344 except for drying at 120°C and feed rate of 6 ml/min, when a too high feed rate disturbed
345 obtaining any dry product. The collected SLM powders were weighed, in order to determine
346 the process yield (according to Eq. 1) and they were subjected to further tests. The results
347 were presented in the form of 3D surface charts showing the relationships between measured
348 values and process parameters (Fig. 1). Evaluated formulation properties and instrument
349 (spray dryer) related parameters were dependent and independent variables, respectively.

350 The obtained SLM powders were characterized by flowability and yield, which
351 allowed for further analysis. The mean d_{50} value in all powders (except the formulation dried
352 at 110°C and the feed rate of 1.2 ml/min) was $14.8 \pm 5.1 \mu\text{m}$ and the percentage of particles
353 $<50 \mu\text{m}$ exceeded $90.2 \pm 7.9\%$. It can be considered that this technique is perfectly suitable
354 for obtaining SLM powder with desirable properties, i.e. small particle size with low span (2.9
355 $\pm 22\%$). An example of the particle size distribution in primary dispersion and in the powder
356 obtained as a result of spray drying (measurement carried out wet and dry, respectively) is
357 presented in Fig. 2. The difference in the measured particle sizes is due to the addition of PVP
358 to the SLM dispersion, which promotes particle sticking during drying (reversible process).
359 Since the measurement was carried out using the dry method (section 2.4.), PVP did not
360 dissolve (as during redispersion in water), thus preventing the measurement of the size of
361 individual lipospheres. In addition, the dry particles tend to agglomerate due to the sticky
362 properties of the lipids and their tendency to adhesion. This does not mean, however, that the
363 primary particle size increases as a result of melting during drying. The preservation of the
364 initial size and shape of the lipid microspheres is confirmed by microscopic observations. Not
365 only the images from the optical microscope, but above all from the high-resolution scanning
366 electron microscope, confirm the unchanged form of the lipid microparticles after the spray
367 drying process (sample microscopic images are presented in Fig. 2). According to the device
368 (Beckman-Coulter LS 13 320) manufacturer's data, the system disperses coherent dry
369 powders without grinding delicate materials. However, as it results from the comparison of
370 the particle sizes measured in the dispersion and after the spray-drying process with the
371 microscopic images and the morphology of the microspheres, the automatic scattering method
372 is insufficient. This is despite the fact that the dry measurement takes place without any
373 disturbances, for a minimum 10 second at controlled obscuration and without operator
374 intervention. On the basis of agglomerates observed on microscopic images of SLM powders
375 (Fig. 2b), in which single particles with preserved morphology were visible, without lumps,

376 without particles fused together or other fragments indicating the melting of the lipid matrix
377 (Fig. 2B), agglomerates were found as the reason for shifting the particle size distribution
378 towards the higher values observed in dry powders. It was found that the obtained results of
379 particle size measurements by dry laser diffraction are similarly affected by agglomerates
380 formed in powders in all formulations, therefore this method can be used to compare their
381 properties. Due to the size of the measured particles, it is still the most accurate method that
382 can be used.

383 The lowest variability depending on the inlet temperature and the feed rate was
384 presented by span and the percentage of particles $<50\ \mu\text{m}$ (Fig. 1 A, B). Obtaining the SLM
385 powders with similar properties in such a wide range of drying conditions means that, taking
386 into account only these two variables, it is possible to dry the aqueous SLM dispersions in
387 almost the entire tested range of parameters, without significantly affecting the properties
388 crucial from the point of view of the dosage form. However, due to the economy of the
389 process, the use of spray drying method should be justified not only by the optimal properties
390 of the products, but also by the efficiency of the process. Determination of percentage yield is
391 very important in the selection of the best parameters of spray drying. In the spray drying
392 process yield of $>45\%$ is considered as acceptable [31]. The low yield in spray drying is
393 mainly due to the small (laboratory) scale of the process. On a large scale, the yield will be
394 greater. Losses during spray drying are usually caused by drying conditions leading to the
395 deposition of droplet or already dried material on the walls of the drying column and cyclone
396 or discharge of fines with the exhaust gas [32]. The yield of the process can be increased with
397 increasing temperature. Unfortunately, too high temperature is not conducive to drying the
398 lipid particle dispersion, which in our case also makes it difficult to achieve optimal yield. In
399 our studies the process yield was a feature that presented the greatest variability depending on
400 the inlet temperature and the feed rate (Fig. 1C). Thus, it is a factor that significantly
401 differentiates formulations depending on the set drying parameters. At the same time,
402 attention should be paid to the fact that the batch size has a large impact on the final yield of
403 the process. The larger the batches, the easier it is to obtain higher process efficiency. The
404 described experiments were carried out on a relatively small batches of 200 g or 300 g (drug-
405 loaded and *placebo* formulations, respectively). However, the main goal was to check the
406 influence of the tested parameters on changes in the drying process yield, and not the absolute
407 amount of recovery, which would have been higher if the process had been conducted on a
408 larger scale.

409 Another common technique that allows drying is the freeze-drying process. In contrast
410 to freeze drying (which the form of the resulting product is “lyophilized cake”) applied to
411 lipid microspheres/nanospheres, spray drying gives the possibility to obtain an already
412 finished, free flowing product, which is required for some applications (e.g. for capsules,
413 sachets, inhalation etc.). In line with our experience, it is easier to disperse spray-dried SLM-
414 powder than freeze-dried SLM-powder, due to the stress associated with freezing and
415 subsequent dehydration of the SLM formulation. The freeze-drying process is also much more
416 expensive and time-consuming, as already mentioned in the introduction, therefore spray
417 drying is often chosen as a faster and more productive process, despite the lower yield
418 [33]. The outlet temperature is a parameter which value depends primarily on the inlet
419 temperature and feed rate [27]. Figure 1D shows how the changes of outlet temperature
420 depended indeed on both the inlet temperature and the feed rate in our experiment. Therefore,
421 the use of outlet temperature as a parameter not only for spray drying control, but also as a
422 criterion for optimizing the entire process was considered. An increase in the inlet
423 temperature from 90°C to 100°C resulted in an increase in the outlet temperature of about
424 4°C, regardless of the feed rate (Fig. 3A), while in the range of inlet temperature from 100°C
425 to 110°C, there were almost no changes in outlet temperature (average change by about 1°C).
426 That proves the outlet temperature stability in this range (Fig. 3B). The greatest increases in
427 outlet temperature (even by about 8°C) were observed when increasing the inlet temperature
428 above 110°C (Fig. 3 A, B). Taking into account the above relations inlet, not outlet
429 temperature was finally considered more appropriate to assess the impact of process
430 parameters on powder properties.

431 From a technological point of view, properties such as powder flow may be of a great
432 importance, especially if the powder is intended to be used as a final form, because in this
433 case, one of the required steps will be to dispense the powder. The dosing process strongly
434 depends on the flowability properties of the final SLM powders. Therefore, to assess the flow
435 properties of obtained dry powders standard pharmacopoeial methods were applied. The
436 results of tests (angle of repose and HR) are presented in Table 3, and their variability,
437 depending on the inlet temperature and feed rate, is also presented in Fig. 1G, H.

438 The flowability expressed as flow rate through an orifice could not be determined,
439 because the obtained SLM formulations were low-flowable powders. Although no
440 macroscopic agglomerates were formed, the adhesive strength between the powder particles
441 was significant. The association of the difficulty of measuring with the surface properties of
442 the powder particles rather than the formation of agglomerates is also confirmed by the fact

443 that it was possible to disperse the powder particles during the particle size measurement (dry
444 measurement technique). Since it was difficult to obtain free flow of the test dry powder
445 through the funnel (10 or 15 mm opening), mechanical shaking was required to obtain the
446 angle of repose cone. Consequently, the obtained cone was irregular, and therefore relatively
447 large variations were noted with some formulations. According to the pharmacopoeial
448 classification, the tested SLM powders could be classified into various groups: from excellent
449 flowing to poor flowing (Table 3), while better properties (lower angle of repose) showed
450 powders dried at lower inlet temperatures (the influence of the flow rate was inconclusive).
451 Hausner ratio, calculated from the bulk and tapped volumes, allowed to classify powders into
452 two categories: as fairly well or passable flowing powders (Table 3), (results slightly different
453 from the results of the angle of repose). Although the angle of repose test showed very good
454 properties of some SLM powders, HR did not fully confirm these results. In our opinion, due
455 to the difficulties described above with measuring the angle of repose and thus the significant
456 differences between the formulations (Table 3), the HR results should be considered as more
457 reliable and representative. Although the HR results indicate slightly worse flow properties of
458 SLM powders (generally fine powders of lipids might present inferior flowability), they still
459 should be considered satisfactory. The European Pharmacopoeia (2.9.36. Powder flow)
460 directly indicates that formulations with an angle of repose in the range of 40-50 degrees are
461 manufactured satisfactorily, and only when an angle of repose exceed 50 degrees, the flow is
462 rarely acceptable for manufacturing purpose (the mean angle of repose of the SLM powders
463 with Compritol is 38.3 ± 8.5 degrees, neither formulation has an angle of repose greater than
464 50 degrees). In the case of the production process, the proper flowability of the tested
465 formulations should be ensured by the addition of lubricating substances.

466 A positive effect on powder flowability might also have lower amount of water, as it
467 can eliminate particle cohesive, tendency to form agglomerates and electrostatic charges.
468 Most studies observe a decrease in powder flow with increasing moisture [34]. Although there
469 is no a specific pharmacopoeial requirements or limits for residual moisture, its level should
470 be determined also taking into account the individual properties of API and other components
471 in dosage form. Moisture content determined in our finally obtained powders was
472 significantly influenced by the feed rate (Fig. 1E). In the SLM powders dried at the higher
473 feed rate (4.5 ml/min), the moisture content exceeded 5% over the entire range of inlet
474 temperatures used (90-120°C). Lower moisture level (about 2.5-3.5%) was observed in
475 powders dried at lower feed rate (1.2-2.4 ml/min). There was no clear correlation between

476 moisture content in the powder and its angle of repose, or moisture and HR. Nevertheless, the
 477 low water content was considered to be a desirable feature of the resulting powders.

478

479 **Table 3**

480 Properties of tested *placebo*-SLM (F1, F2) and API-loaded SLM dry powders depending on
 481 the conditions of spray drying, classification based on pharmacopoeial criteria (chapter 2.9.36.
 482 Powder flow, European Pharmacopoeia)

Conditions of the drying process			Values of tested parameters		
Inlet temperature [°C]	Feed rate [ml/min]	Angle of repose [°] ± SD	Flow property	Hausner ratio	Flow character
<i>SLM placebo</i> with Compritol					
90	1.2	45.8 ± 4.1	Passable	1.24	Fair
90	2.4	26.5 ± 1.6	Excellent	1.33	Passable
90	4.5	30.7 ± 1.7	Excellent	1.23	Fair
100	1.2	31.7 ± 0.6	Good	1.29	Passable
100	2.4	37.9 ± 1.1	Fair	1.19	Fair
100	4.5	46.2 ± 2.9	Poor	1.22	Fair
110	1.2	27.4 ± 0.3	Excellent	1.32	Passable
110	2.4	43.2 ± 1.0	Passable	1.26	Passable
110	4.5	47.3 ± 3.2	Poor	1.28	Passable
110	6.0	n.t.	n.t.	n.t.	n.t.
120	1.2	n.t.	n.t.	n.t.	n.t.
120	4.5	46.6 ± 1.2	Poor	1.24	Fair
<i>SLM placebo</i> with stearic acid					
75	3	34.6 ± 1.2	Good	1.23	Fair
80	2.4	35.2 ± 0.7	Good	1.21	Fair
80	3	38.0 ± 0.9	Fair	1.17	Good
API-loaded SLM with Compritol spray-dried under inlet temp. 90°C and feed rate 2.4 ml/min					
Formulation F3		29.9 ± 0.3	Excellent	1.32	Passable
Formulation F4		37.6 ± 1.2	Fair	1.30	Passable
Formulation F7		34.0 ± 1.1	Good	1.33	Passable
Formulation F8		31.5 ± 1.0	Good	1.32	Passable

483 n.t. – not tested

484

485 When considering the optimal powder properties, obtained formulations should be
 486 characterized by the largest percentage of particles with a size below 50 µm, the smallest
 487 span, with the best flow properties, low moisture content and with maximum yield at the same
 488 time (Fig. 1). The above criteria are best met by SLMs dried in conditions of both low inlet
 489 temperatures and low feed rates. In the tested range of parameter values, taking into account

490 the properties of the final powders, the optimal parameters for spray drying of the aqueous
491 dispersion of SLM with Compritol were: 90°C inlet temperature and feed rate 2.4 ml/min
492 (which corresponds to outlet temperature about 43°C). SLM powders dried under such
493 conditions characterized by the following properties: excellent angle of repose (26.5°),
494 passable value of HR (1.33), low moisture content – 3%, satisfactory yield – 59%, span – 2.5
495 and the percentage of particles <50 µm – 95%.

496

497 3.3. Spray drying of aqueous dispersions of stearic acid-SLM

498 Similar to the formulations with Compritol, SLM dispersions with stearic acid were
499 spray dried in the range of process parameters indicated in Table 2. Although the melting
500 point of stearic acid is similar to Compritol (69,6°C for stearic acid and 69-74°C for glyceryl
501 behenate), more difficulties were observed when spray drying microspheres with stearic acid
502 than with Compritol. Even slight and short-term destabilization of parameters during drying
503 step resulted in melting of lipid particles, sticking to the elements of the dryer (mainly cyclon)
504 and failure of the whole process. When drying SLM dispersions with stearic acid using
505 different conditions, two batches process at the highest temperature (90°C and 100°C) and
506 with the lowest feed rate were completely unsuccessful (no yield). All other dried
507 formulations were weighed to determine yield and then, they were further tested as already
508 described for Compritol powders. The collected data and test results were compiled in the
509 form of 3D surface charts (Fig. 4) showing the relationships between powder properties and
510 drying process parameters.

511 In diagrams (Fig. 4A and B), showing the relationship between span or particle size
512 and drying parameters, for powders with stearic acid, greater variation was observed than in
513 SLM with Compritol, and thus the area of obtaining favorable properties is narrowed down,
514 mainly to the range of lower temperatures, with less influence of the feed rate. Similar to
515 formulations with Compritol, in spray dried SLM powders with stearic acid the greatest
516 variability depending on the inlet temperature and the feed rate was presented by the process
517 yield (Fig. 4C). As indicated by Fig. 4E the residual moisture in powders with stearic acid was
518 not as varied and dependent on the feed rate as in the case of SLM with Compritol, and its
519 highest range was coincided, oddly enough, with the highest drying temperatures. The chart
520 of outlet temperature dependence on inlet temperature and feed rate has a similar shape,
521 regardless of the lipid forming the matrix of the microspheres (Fig. 1D vs. Fig. 4D).

522 The flowability of SLM with stearic acid were determined only for selected powders
523 with the best properties, spray dried at 75-80°C inlet temperature with the feed rate 2.4 or 3
524 ml/min. The results of both: the angle of repose and the Hausner ratio classified the tested
525 formulations between good and fairly good flowability (Table 3).

526 When considering the best powder properties, obtained dry powders with stearic acid
527 should meet the same requirements as for SLM with Compritol. This was possible, as in SLM
528 with Compritol, when the dispersions were dried under conditions of both low inlet
529 temperatures and low feed rates (among the values tested for stearic acid).

530 In conclusion, the optimal spray drying parameters for SLM aqueous dispersion with
531 stearic acid were: 80°C inlet temperature with feed rate 3.0 ml/min (which corresponds to
532 outlet temperature about 36°C). SLM powders dried under such conditions characterized by
533 the following properties: fair angle of repose (38°), good value of HR (1.17), low moisture
534 content – 3.5%, yield – 48%, span – 2.5 and the percentage of particles <50 µm – 98%.

535

536 3.4. Optimizing the spray drying process of SLM dispersions

537 All the obtained results from tested *placebo* SLM formulations with different lipids
538 were collected in the form of three-dimensional surface charts and were also subjected to
539 statistical analysis. The conducted experiments allowed to indicate the critical factors and the
540 best process conditions. In accordance with the adopted assumptions obtained dry SLM
541 powder should fulfill the following requirements: good flowability, percentage share of
542 particles <50 µm (at least 90%), span (as small as possible), acceptable yield (at least 50%),
543 and moisture content (not more than 3%).

544 Appropriate ranges of process parameters, allowing to obtain a product with the
545 indicated characteristics, can be confirmed by visual inspection of 3D charts. The analysis of
546 the charts of preparations with Compritol (Fig. 1) and stearic acid (Fig. 4) also allows to
547 verify the potential effects of changing the tested independent variables (process parameters)
548 on the properties of the formulation.

549 Although the technology of the spray-drying process eliminates significant exposure
550 of the spray-dried dispersion to elevated temperature, only a process that does not melt the
551 lipid from the matrix of microspheres can be considered correctly carried out. It depends on
552 the proper adjustment of the process input parameters (inlet temperature, feed rate).

553 Performed experiments showed that only selected combinations of tested parameters
554 (inlet temperature and feed rate) resulted in obtaining final product with desired properties. In

555 Table 4, the various configurations of the monitored process parameters were color-coded. In
 556 the case of SLM with Compritol, the indicated range of parameters (yellow) is even quite
 557 wide (in the case of SLM with stearic acid, it is much more limited).

558

559 **Table 4**

560 Categorization of the conditions of the spray drying process depending on the properties of
 561 the obtained powder and the course of the process with the simultaneous differentiation of the
 562 lipid forming the SLM matrix.

SLM placebo formulations with Compritol (F1)														
Inlet temp. [°C]			90	90	90	100	100	100	110	110	110	120	120	
Feed rate [ml/min]	n.t.		1.2	2.4	4.5	1.2	2.4	4.5	1.2	2.4	4.5	6.0	1.2	4.5
Outlet temp. [°C]			48	43	36	52	46	41	54	46	42	32	62	49
SLM placebo formulations with stearic acid (F2)														
Inlet temp. [°C]	75	80	80	80	n.t.	90	90	n.t.	100	100				
Feed rate [ml/min]	3.0	2.4	3.0	4.5	n.t.	2.4	3.0	n.t.	2.4	4.5			n.t.	
Outlet temp. [°C]	33	40	36	32		44	40		50	42				

563 n.t. – not tested, green color – optimal drying conditions, yellow color – acceptable drying conditions, red color –
 564 unfavorable drying conditions

565

566 During the drying of SLM with stearic acid in conditions other than those marked in
 567 green, there were fluctuations in the outlet temperature affecting the process (an increase in
 568 the outlet temperature sometimes caused a further increase in this temperature, and
 569 consequently destabilization of the entire process). The smooth course of the drying process
 570 was disturbed mainly by the deposition of a part of the dried powder in the cyclone (red in
 571 Table 4). This phenomenon could be due to partial lipid melting during drying, as this
 572 occurred mainly at higher inlet temperatures with lower feed rates (Table 4). SLMs with
 573 stearic acid were particularly susceptible to this phenomenon. Consequently, less product
 574 ended up in the dry particles collector, resulting in reduced process efficiency. In extreme
 575 situations, the drying gas flow was even obstructed and the cyclone was clogged. Moreover,
 576 in some experiments, in the final product collector single lipid lumps were observed.

577 Analyzing surface charts, it was noticed that to prevent an adverse increase in outlet
 578 temperature, it is advisable that when raising the drying temperature (inlet temperature) feed
 579 rate should be also increased. During optimization of the conditions of the spray drying
 580 process of SLM with stearic acid it turned out, that despite the inlet temperature reduction

581 (from 90°C to 80°C), the feed rate also need to be increased (from 2.4 to 3.0 ml/min). In
582 addition, to reduce the risk of outlet temperature increasing during drying when the feed rate
583 is lower, conditions with higher feed rate were marked as more favorable (green color in
584 Table 4), although the use of both parameters combinations is possible.

585 Comparing the behavior of SLM depending on the lipid forming the matrix of
586 microspheres, stearic acid formulations proved to be more demanding during processing.
587 Paradoxically, despite a similar melting point, a greater sensitivity of SLM with stearic acid
588 than with Compritol was observed to the already described partial melting and cyclone
589 clogging. Moreover, despite the use of a lower inlet temperature, a sufficiently high feed rate
590 was important (Table 4). Ultimately, the parameters that allowed to obtain powders with the
591 best properties require drying SLM with stearic acid under slightly milder conditions (lower
592 inlet temperature and at the same time higher feed rate) compared to SML containing
593 Compritol.

594 In summary, the choice of drying conditions for SLM with stearic acid is much less
595 flexible compared to SLM with Compritol. Thus, the type of lipid used in SLM is crucial for
596 the properties of SLM powder and can significantly affect the drying process itself.

597

598 3.5. Spray drying of drug-loaded SLM dispersions

599 To assess the influence of the active substances incorporation in the carrier on the
600 drying process and the properties of the powder obtained, SLM formulations with model drug
601 substances (CsA and SPIR, Table 1) were spray dried using the process conditions optimized
602 for *placebo* SLM. This publication focuses solely on the possibility of API-loaded SLM spray
603 drying using conditions that have been established and optimized using a *placebo*
604 formulation. Therefore, dry SLM powders with API were characterized only in the same way
605 as *placebo* formulations (particle size, yield, flowability, etc.). Important biopharmaceutical
606 aspects, such as the influence of the spray-drying process on the release rate of the active
607 substance or the distribution of API within the lipid microspheres, including drug substance
608 expulsion from the lipid matrix, assessed by various instrumental techniques, are described in
609 a separate paper [30].

610 API loaded dispersions with Compritol as a lipid matrix (F3-F4 with 0.1 and 1.0% of
611 CsA, respectively as well as F7-F8 with 0.1 and 0.5% of SPIR, respectively) were drying at
612 inlet temperature 90°C with feed spray rate 2.4 ml/min, when dispersions with stearic acid as
613 a lipid matrix (F5-F6 with 0.1 and 1.0% of CsA, respectively as well as F9-F10 with 0.1 and

614 0.5% of SPIR, respectively) were drying at inlet temperature 80°C and feed rate 3.0 ml/min.
 615 All SLM formulations with API were successfully spray dried using conditions selected as the
 616 optimal in the *placebo* formulations studies. In each case, the process ran smoothly and did
 617 not require any modification, regardless of the type and concentration of API. Obtained SLM
 618 powders with API mostly met all the criteria for the obtained dry products, assumed during
 619 optimization of the drying process parameters for *placebo* formulations. In Table 5 the yields
 620 and moisture residuals values of dry powders with CsA or SPIR are summarized.

621

622 **Table 5**

623 Characteristics of spray dried SLM powders with API

Investigated powder properties	Formulations with Compritol				Formulations with stearic acid			
	F3	F4	F7	F8	F5	F6	F9	F10
Yield (%)	48.0%	64.0%	52.6%	53.0%	50.1%	50.5%	47.6%	37.2%
Moisture content (%)	3.4%	3.2%	3.4%	2.6%	3.2%	2.8%	3.3%	4.1%

624

625

626 The F10 powder was a formulation which properties deviated the most from the
 627 accepted values. All other formulations can be considered optimally spray dried. Other
 628 criteria (span and percentage of particles below 50 μm) were also met. In Fig. 5 the particle
 629 size distributions of spray dried powder of two selected formulations (F4 – SLM with CsA
 630 and Compritol, F5 – SLM with CsA and stearic acid) were presented. The mean d_{50} value in
 631 API-loaded powders was 13 μm and 11 μm , while the percentage of particles $<50 \mu\text{m}$
 632 exceeded 95% and 98% for F4 and F5 formulations, respectively.

633 Moreover, tested powders demonstrated good flowability properties, as evidenced by
 634 the satisfactory values of the angle of repose, as well as the HR values, similar to the *placebo*
 635 formulations (Table 3). When the dry powders were redispersed in water, no precipitation of
 636 the active substance was observed in the microscopic image. In the previous work, no
 637 significant changes in API distribution as a result of the spray drying process were found. A
 638 detailed description of the research on spray-dried aqueous SLM dispersions loaded with API
 639 can be found in a separate publication [30].

640 Summing up the results of drying SLM dispersions with API, it can be concluded that
 641 the introduction of the drug substance (each of the two chosen as a model drug) into the
 642 formulation, even at a fairly significant concentration (1% in the dispersion, which

643 corresponds to 10% relative to the amount of lipid) did not have a significant influence on the
644 drying process.

645 The results evidence that process parameters developed for *placebo* SLM formulations
646 can be effectively used when drying SLM with different API, also in different concentrations,
647 if the composition contains the same matrix-forming lipid.

648 The properties of obtained SLM powders with CsA or SPIR allow for their further use
649 and administration in dry form or after prior reconstitution to a liquid aqueous dispersion
650 (when the dry powders were redispersed in water, no precipitation of the API was observed in
651 the microscopic image.). From a practical point of view, it is of a great important, that it is
652 possible, especially at least on a laboratory scale, to optimize the spray drying process of
653 selected formulation with API using *placebo* formulation with the same lipid forming the
654 microparticles matrix, thus saving often expensive and available in small quantities active
655 substances.

656

657 **4. Conclusions**

658 SLM dispersions, both *placebo* and with API (e.g. CsA and SPIR in various
659 concentrations), can be effectively converted from aqueous dispersion to dry powder by spray
660 drying technique. The conducted studies allowed us to optimize spray drying process of
661 aqueous SLM dispersions containing two different lipids (Compritol and stearic acid) as
662 microspheres matrix. Performed experiments showed that only selected combinations of
663 tested parameters (inlet temperature and feed rate) resulted in obtaining final product with
664 desired properties.

665 The use of 3D surface charts, developed on the basis of the results of experiments
666 carried out with different values of independent variables, allows to predict the values of
667 dependent variables depending on the conditions of the process. Flexibility in the selection of
668 drying conditions (inlet temperature and feed rate) depends primarily on the lipid forming the
669 matrix of microspheres. The choice of drying conditions for SLM with stearic acid is much
670 less flexible compared to SLM with Compritol. Thus the lipid forming the matrix of
671 microspheres is the basic factor for which the appropriate drying parameters must be selected.
672 The results of conducted experiments and further statistical analysis of the obtained results as
673 the most favorable conditions for conducting the spray drying process of SLM dispersions
674 with Compritol indicated the inlet temperature of 90°C and the feed spray rate 2.4 ml/min,
675 while for SLM with stearic acid inlet temperature was 80°C with feed spray rate 3 ml/min.

676 Spray drying can be carried out completely without the use of organic solvents due to
677 the fact that the aqueous dispersion of lipid microspheres is dried. In addition, it is possible to
678 optimize the drying process on *placebo* formulations to conditions that will then be
679 successfully used for drying SLM with API.

680

681 **Declaration of interest**

682 The author reports no conflict of interest.

683

684 **References**

685

686 [1] M. Lengyel, N. Kállai-Szabó, V. Antal, A.J. Laki, I. Antal, Microparticles,
687 microspheres, and microcapsules for advanced drug delivery, *Sci. Pharm.* 87 (2019).
688 doi:10.3390/scipharm87030020.

689 [2] P. Yalavarthi, T. Dudala, N. Mudumala, V. Pasupati, J. Thanniru, H. Vadlamudi, G.
690 Yaga, A perspective overview on lipospheres as lipid carrier systems, *Int. J. Pharm.*
691 *Investig.* 4 (2014) 149. doi:10.4103/2230-973x.143112.

692 [3] S. Jaspert, G. Piel, L. Delattre, B. Evrard, Solid lipid microparticles: formulation,
693 preparation, characterisation, drug release and applications., *Expert Opin. Drug Deliv.*
694 2 (2005) 75–87. doi:10.1517/17425247.2.1.75.

695 [4] L. Battaglia, M. Gallarate, P.P. Panciani, E. Ugazio, S. Sapino, E. Peira, D. Chirio,
696 Techniques for the Preparation of Solid Lipid Nano and Microparticles, *Appl.*
697 *Nanotechnol. Drug Deliv.* (2014). doi:10.5772/58405.

698 [5] S. Han, P. Dwivedi, F.A. Mangrio, M. Dwivedi, R. Khatik, D.E. Cohn, T. Si, R.X. Xu,
699 Sustained release paclitaxel-loaded core-shell-structured solid lipid microparticles for
700 intraperitoneal chemotherapy of ovarian cancer, *Artif. Cells, Nanomedicine*
701 *Biotechnol.* 47 (2019) 957–967. doi:10.1080/21691401.2019.1576705.

702 [6] C. Wu, X. Luo, S.G. Baldursdottir, M. Yang, X. Sun, H. Mu, In vivo evaluation of
703 solid lipid microparticles and hybrid polymer-lipid microparticles for sustained
704 delivery of leuprolide, *Eur. J. Pharm. Biopharm.* 142 (2019) 315–321.
705 doi:10.1016/j.ejpb.2019.07.010.

706 [7] P.C.H. Wong, P.W.S. Heng, L.W. Chan, Spray congealing as a microencapsulation
707 technique to develop modified-release ibuprofen solid lipid microparticles: The effect
708 of matrix type, polymeric additives and drug-matrix miscibility, *J. Microencapsul.* 32
709 (2015) 725–736. doi:10.3109/02652048.2015.1073387.

710 [8] E. Wolska, M. Sznitowska, Technology of stable, prolonged-release eye-drops
711 containing Cyclosporine A, distributed between lipid matrix and surface of the solid
712 lipid microspheres (SLM), *Int. J. Pharm.* 441 (2013).
713 doi:10.1016/j.ijpharm.2012.11.009.

714 [9] K. Jores, A. Haberland, S. Wartewig, K. Mäder, W. Mehnert, Solid Lipid

- 715 Nanoparticles (SLN) and oil-loaded SLN studied by spectrofluorometry and raman
716 spectroscopy, *Pharm. Res.* 22 (2005) 1887–1897. doi:10.1007/s11095-005-7148-5.
- 717 [10] V. Mishra, K.K. Bansal, A. Verma, N. Yadav, S. Thakur, K. Sudhakar, J.M.
718 Rosenholm, Solid lipid nanoparticles: Emerging colloidal nano drug delivery systems,
719 *Pharmaceutics*. 10 (2018) 1–21. doi:10.3390/pharmaceutics10040191.
- 720 [11] M. Sznitowska, E. Wolska, H. Baranska, K. Cal, J. Pietkiewicz, The effect of a lipid
721 composition and a surfactant on the characteristics of the solid lipid microspheres and
722 nanospheres (SLM and SLN), *Eur. J. Pharm. Biopharm.* 110 (2017).
723 doi:10.1016/j.ejpb.2016.10.023.
- 724 [12] M.A. Momoh, F.C. Kenekwukwu, A.A. Attama, Formulation and evaluation of novel
725 solid lipid microparticles as a sustained release system for the delivery of metformin
726 hydrochloride, *Drug Deliv.* 20 (2013) 102–111. doi:10.3109/10717544.2013.779329.
- 727 [13] S. Bertoni, B. Albertini, C. Facchini, C. Prata, N. Passerini, Glutathione-loaded solid
728 lipid microparticles as innovative delivery system for oral antioxidant therapy,
729 *Pharmaceutics*. 11 (2019). doi:10.3390/pharmaceutics11080364.
- 730 [14] E.C. Umeyor, F.C. Kenekwukwu, J.D. Ogbonna, S.A. Chime, A. Attama, Preparation
731 of novel solid lipid microparticles loaded with gentamicin and its evaluation in vitro
732 and in vivo, *J. Microencapsul.* 29 (2012) 296–307.
733 doi:10.3109/02652048.2011.651495.
- 734 [15] Y. Rahimpour, Y. Javadzadeh, H. Hamishehkar, Solid lipid microparticles for
735 enhanced dermal delivery of tetracycline HCl, *Colloids Surfaces B Biointerfaces*. 145
736 (2016) 14–20. doi:10.1016/j.colsurfb.2016.04.034.
- 737 [16] E. Wolska, M. Sznitowska, J. Chorążewicz, O. Szerkus, A. Radwańska, M.J.
738 Markuszewski, R. Kaliszan, K. Raczynska, Ocular irritation and cyclosporine A
739 distribution in the eye tissues after administration of Solid Lipid Microparticles in the
740 rabbit model, *Eur. J. Pharm. Sci.* 121 (2018). doi:10.1016/j.ejps.2018.05.015.
- 741 [17] M. Mezzena, S. Scalia, P.M. Young, D. Traini, Solid lipid budesonide microparticles
742 for controlled release inhalation therapy, *AAPS J.* 11 (2009) 771–778.
743 doi:10.1208/s12248-009-9148-6.
- 744 [18] Y.Z. Li, X. Sun, T. Gong, J. Liu, J. Zuo, Z.R. Zhang, Inhalable microparticles as
745 carriers for pulmonary delivery of thymopentin-loaded solid lipid nanoparticles, *Pharm.*
746 *Res.* 27 (2010) 1977–1986. doi:10.1007/s11095-010-0201-z.
- 747 [19] E. Nemati, A. Mokhtarzadeh, V. Panahi-Azar, A. Mohammadi, H. Hamishehkar, M.
748 Mesgari-Abbasi, J. Ezzati Nazhad Dolatabadi, M. de la Guardia, Ethambutol-Loaded
749 Solid Lipid Nanoparticles as Dry Powder Inhalable Formulation for Tuberculosis
750 Therapy, *AAPS PharmSciTech.* 20 (2019) 1–9. doi:10.1208/s12249-019-1334-y.
- 751 [20] M.E. Ali, A. Lamprecht, Spray freeze drying as an alternative technique for
752 lyophilization of polymeric and lipid-based nanoparticles, *Int. J. Pharm.* 516 (2017)
753 170–177. doi:10.1016/j.ijpharm.2016.11.023.
- 754 [21] C. Freitas, R.H. Müller, Spray-drying of solid lipid nanoparticles (SLN(TM)), *Eur. J.*
755 *Pharm. Biopharm.* 46 (1998) 145–151. doi:10.1016/S0939-6411(97)00172-0.

- 756 [22] C. Freitas, R.H. Müller, Correlation between long-term stability of solid lipid
757 nanoparticles (SLNTM) and crystallinity of the lipid phase, *Eur. J. Pharm. Biopharm.*
758 47 (1999) 125–132. doi:10.1016/S0939-6411(98)00074-5.
- 759 [23] S. Martins, I. Thon, D.C. Ferreira, E.B. Souto, M. Brandl, Physicochemical properties
760 of lipid nanoparticles: effect of lipid and surfactant composition, *Drug Dev. Ind.*
761 *Pharm.* 37 (2011) 815–824. doi:10.3109/03639045.2010.545414.
- 762 [24] B. Albertini, N. Passerini, M.L. González-Rodríguez, B. Perissutti, L. Rodriguez,
763 Effect of Aerosil[®] on the properties of lipid controlled release microparticles, *J.*
764 *Control. Release.* 100 (2004) 233–246. doi:10.1016/j.jconrel.2004.08.013.
- 765 [25] E.A. Fouad, M. El-Badry, G.M. Mahrous, I.A. Alsarra, Z. Alashbban, F.K. Alanazi, In
766 vitro investigation for embedding dextromethorphan in lipids using spray drying, *Dig.*
767 *J. Nanomater. Biostructures.* 6 (2011) 1129–1139.
- 768 [26] C. Wu, M. van de Weert, S.G. Baldursdottir, M. Yang, H. Mu, Effect of excipients on
769 encapsulation and release of insulin from spray-dried solid lipid microparticles, *Int. J.*
770 *Pharm.* 550 (2018) 439–446. doi:10.1016/j.ijpharm.2018.09.007.
- 771 [27] D. Santos, A.C. Mauricio, V. Sencadas, J.D. Santos, M.H. Fernandes, P.S. Gomes,
772 Spray drying: an overview, *IntechOpen.* Chapter 2 (2018) 9–35.
773 doi.org/10.5772/intechopen.72247.
- 774 [28] S. Kumar, J.K. Randhawa, Solid lipid nanoparticles of stearic acid for the drug delivery
775 of paliperidone, *RSC Adv.* 5 (2015) 68743–68750. doi:10.1039/c5ra10642g.
- 776 [29] M.H. Aburahma, S.M. Badr-eldin, Compritol 888 ATO: a multifunctional lipid
777 excipient in drug delivery systems and nanopharmaceuticals, *Expert Opin. Drug Deliv.*
778 (2014) 1–19. doi:10.1517/17425247.2014.935335.
- 779 [30] E. Wolska, M. Sznitowska, K. Krzemińska, M. Ferreira Monteiro, Analytical
780 techniques for the assessment of drug-lipid interactions and the active substance
781 distribution in liquid dispersions of solid lipid microparticles (SLM) produced *de novo*
782 and reconstituted from spray-dried powders. *Pharmaceutics.* 12 (2020) 664–686.
783 doi.org/10.3390/pharmaceutics12070664
- 784 [31] K. Sollohub, K. Cal, Spray drying technique: II. Current applications in pharmaceutical
785 technology, *J. Pharm. Sci.* 99 (2010) 587–597. Doi:10.1002/jps.21963.
- 786 [32] M. Maury, K. Murphy, S. Kumar, L. Shi, G. Lee, Effects of process variables on the
787 powder yield of spray-dried trehalose on a laboratory spray-dryer. *Eur. J. Phar.*
788 *Biopharm.* 59 (2005) 565–573. doi:10.1016/j.ejpb.2004.10.002.
- 789 [33] E.F. Santo, L.K. Lima, A.P. Torres, G. Oliveira, E.G. Ponsano, Comparison between
790 freeze and spray drying to obtain powder *Rubrivivax gelatinosus* biomass. *Food Sci.*
791 *Technol.* 33 (2013) 47–51. dx.doi.org/10.1590/S0101-20612013005000008
- 792 [34] N. Sandler, K. Reiche, J. Heinamaki, J. Yliruusi, Effect of moisture on powder flow
793 properties of theophylline, *Pharmaceutics.* 2 (2010) 275–290.
794 doi:10.3390/pharmaceutics2030275.

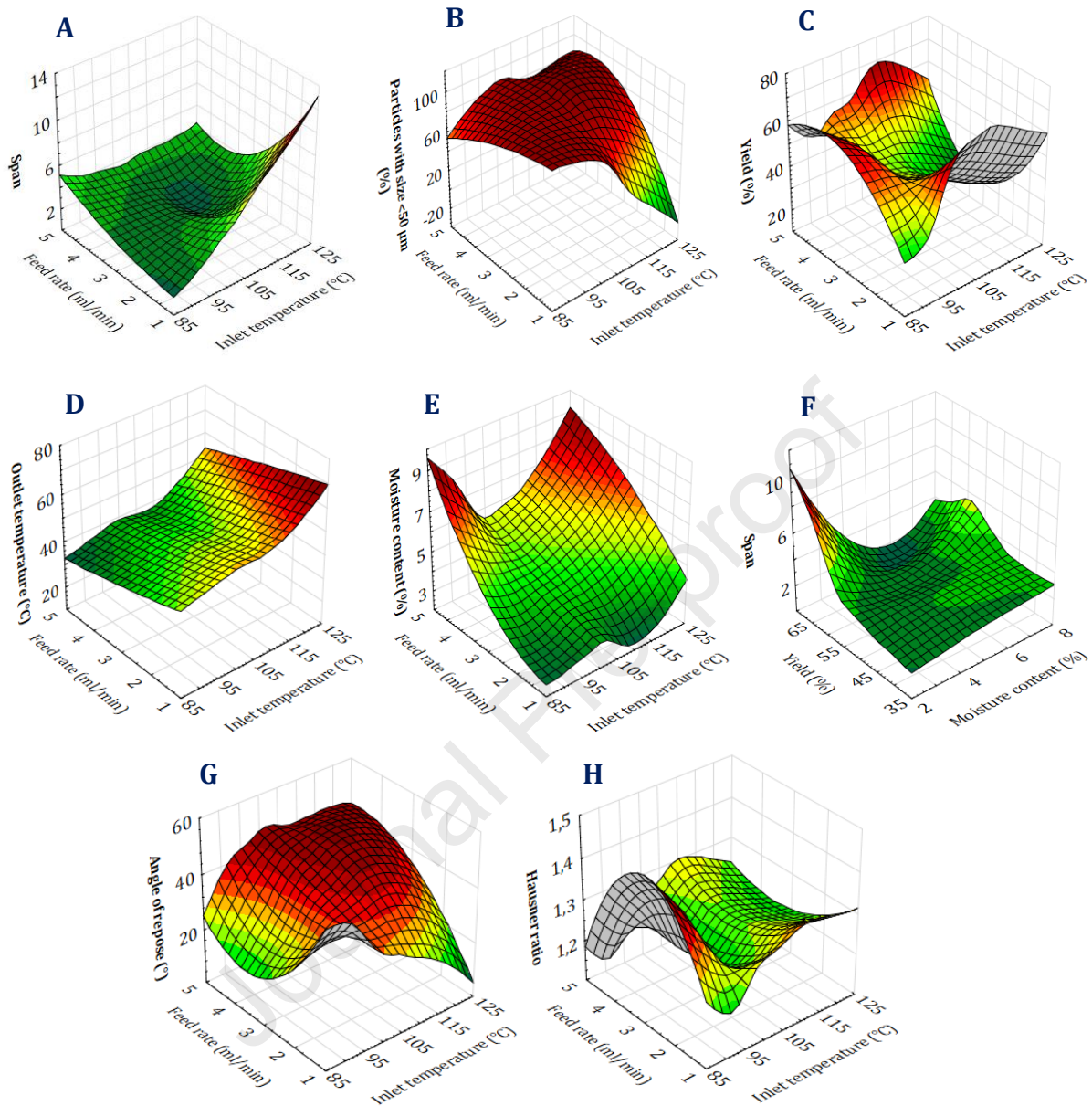


Figure 1. Surface plots 3D presenting the impact of investigated factors on the properties of spray dried SLM powders with Compritol. The dependence of (A) span, (B) particle size <50 μm , (C) yield, (D) outlet temperature, (E) moisture content, (G) angle of repose, (H) Hausner ratio on inlet temperature and feed rate; and dependence of (F) span on yield and moisture content is presented. Brown color corresponds to the highest values and dark green corresponds to the lowest values.

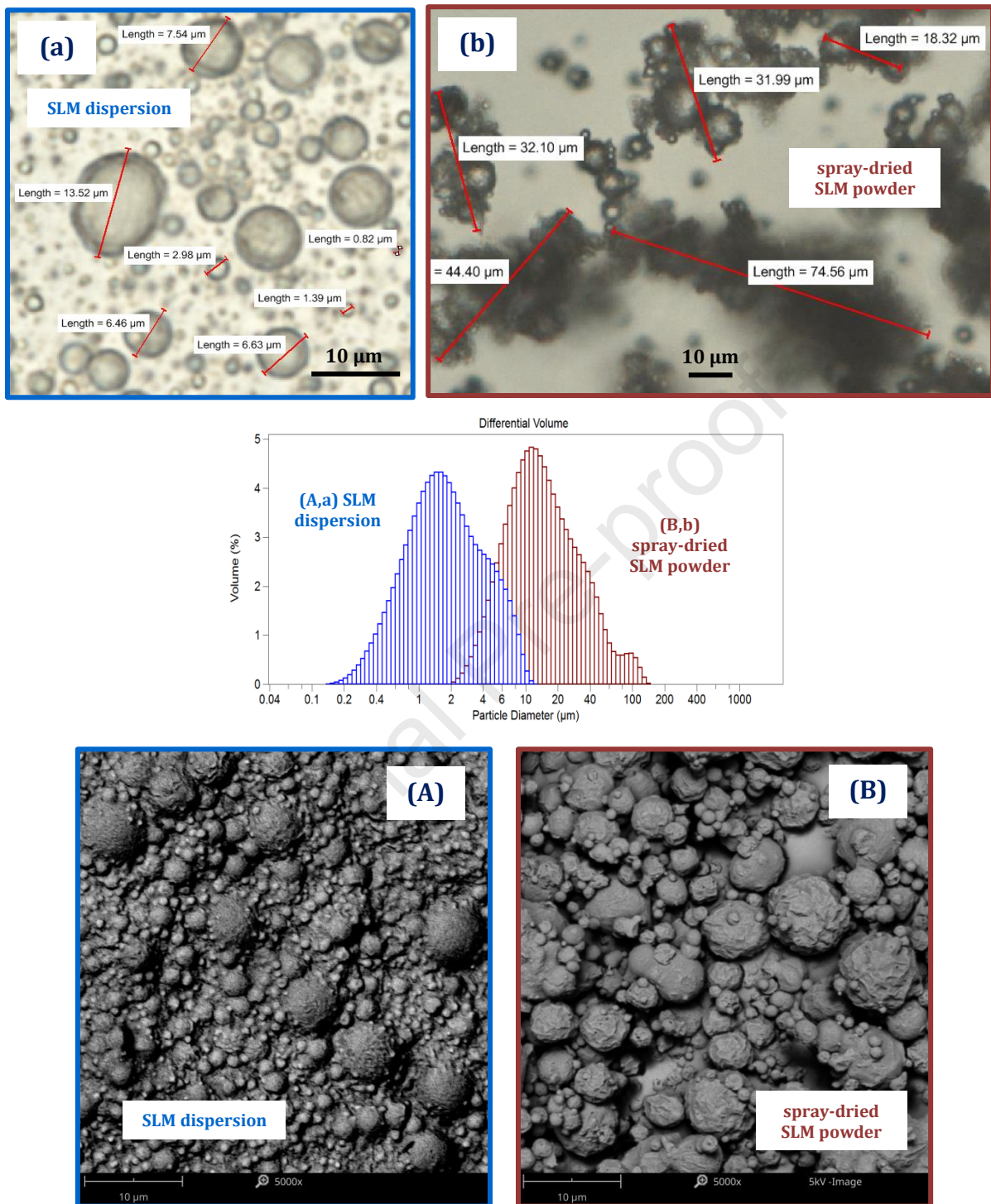


Figure 2. Particle size distribution profiles of selected *placebo* SLM with Compritol: before spray drying (aqueous dispersion) and after spray drying (SLM powder), as well as optical microscopic picture (a, b) and scanning electron micrographs (A, B) of SLM dispersion (A, a) and spray-dried SLM powder (B, b).

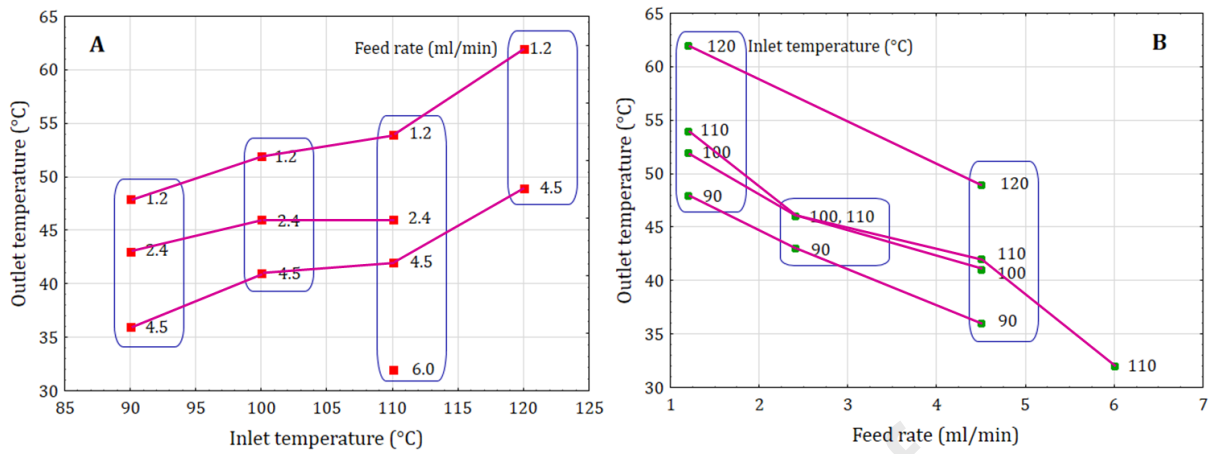


Figure 3. Graphs showing the relationship between (A) inlet temperature, (B) feed rate and outlet temperature.

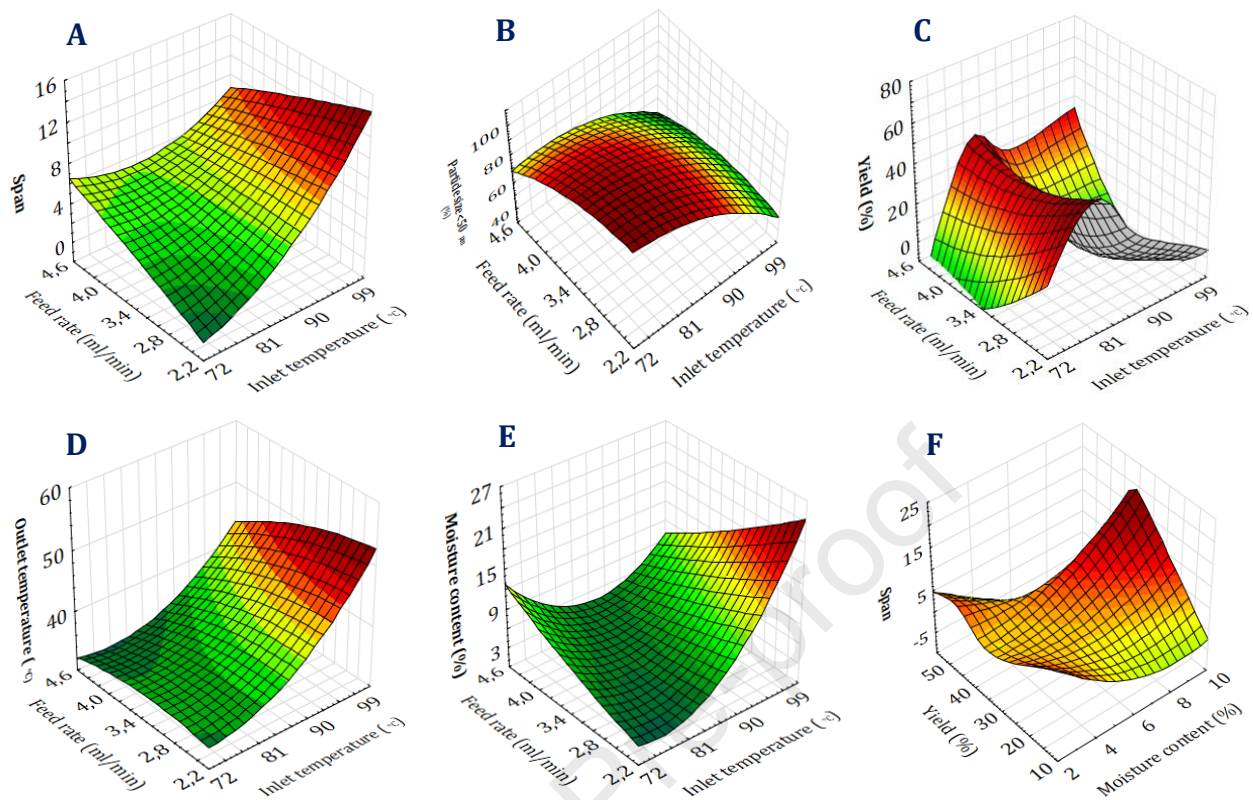


Figure 4. Surface plots 3D presenting the impact of investigated factors on the properties of spray dried SLM powders with stearic acid. The dependence of (A) span, (B) particle size <50 μm , (C) yield, (D) outlet temperature, (E) moisture content on inlet temperature and feed rate; and dependence of (F) span on yield and moisture content is presented. Brown color correspond to the highest values and dark green correspond to the lowest values.

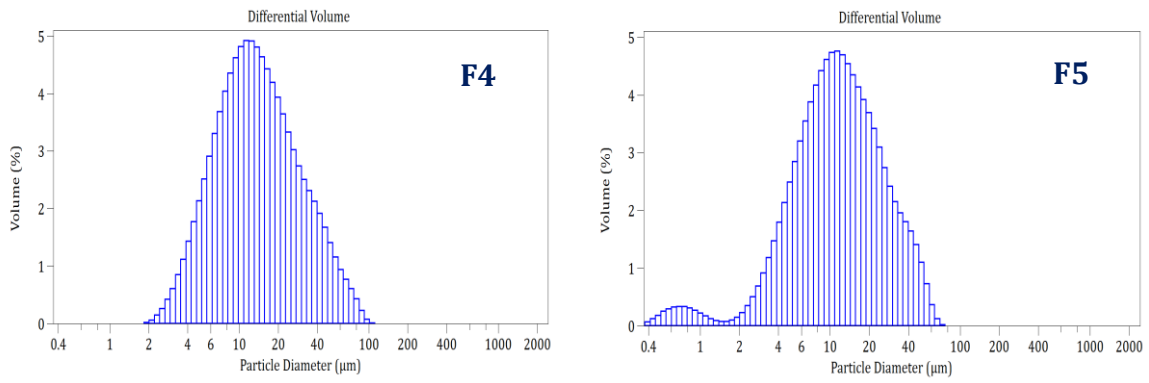
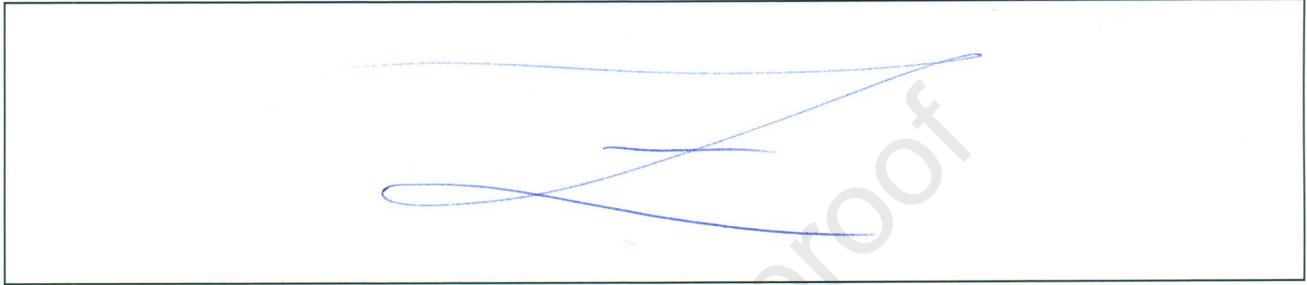


Figure 5. Particle size distribution profiles of selected CsA-loaded spray dried SLM powders.

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: —



E. Wolska

Journal Pre-proof